**Evaluation of the Fungal Species Associated with the Spoilage Of Cocoyam Obtained in Benin City, Nigeria.**

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**ABSTRACT**

A staple food in many cultures around the world, particularly in Nigeria, is cocoyam. However, this food product is vulnerable to pathogen invasion, especially fungus, and post-harvest deterioration. Therefore, the purpose of this study was to assess the fungal species linked to the spoiling of cocoyam purchased in Benin City. From a cocoyam barn in a nearby market, corms with signs of post-harvest rot and corms in good condition were acquired. Spoiled tissues from the corms were removed using a sterile knife. A section of the cut-out area was then infected on PDA plates, which were then incubated at 270C for four days before being checked every day for the growth of fungus. As a control, uninoculated cocoyams without any isolation were used. For fourteen days, the cormels were cultured and monitored. *Mucor cercinelloides, Aspergillus niger, Aspergillus fumigatus, Rhizopus stolonifer, Fusarium oxysporum, and Penicillium cyclopium* were the six fungi that were isolated from the spoiled cormels. The weight loss percentage of the cocoyam samples inoculated with various fungal infections varied from 49.57% to 64.66%, whereas the control sample showed a weight reduction of 3.77 %. With the exception of Penicillium, whose alterations only became apparent on day 8, all isolates showed notable modifications between day 4 and day 9. If these organisms infect the cocoyam, growers and customers may suffer significant financial losses due to the drastic decrease in weight in such a short amount of time. Because cocoyams are a food plant that many people eat, they must be planted, stored, and even cooked with extreme caution. Furthermore, it's critical to take aseptic measures to totally prevent fungal growth, which can endanger consumers.

Keywords: Cocoyam , Fungi isolates, cormels, Colocasia esculenta, Araceae

**INTRODUCTION**

Cocoyam is a perennial member of the Araceae family of herbaceous roots, cocoyams are monocotyledonous. The two genera of cocoyams that are grown for food are Colocasia and Xanthosoma. They are typically made up of a big, spherical corm, or inflated underground storage stem, from which a few enormous leaves sprout. There are two edible species of cocoyam: *Xanthosoma sagittifolium* (L) Schott, popularly known as Tannia, and *C. esculenta*, widely known as Taro. (Lewu *et al*., 2010). Both are typically grown for their larger underground corms and cormels, or tubers, which are rich in starch. The plant is cultivated mostly for its tasty roots, even though all of its components are edible. It is thought that colocasia evolved in the Indo-Malaysian region and then expanded to the Eastern Mediterranean, the Pacific Islands, and ultimately Africa . Native to tropical Central and South America, as well as the Caribbean, *Xanthosoma* spp. have been grown and eaten there since pre-Columbian times, and it was from these regions that it spread to other areas of the world .

*Xanthosoma sagittifolium* is known as ede-oku in Igbo, whereas *Colocasia esculenta* is known as ede-uli. The Yoruba call cocoyam "koko." It is one of the top six root and tuber crops in the world (Agu *et al*., 2016). It is domesticated in most communities in Oceania, Africa, and Asia and is pantropical in nature. West Africa is the region with the greatest area under cultivation and, thus, the highest output volume. With an annual production of 4.55 million metric tonnes in 2012, Nigeria topped the list of countries producing cocoyams; this amounted to 61.2 and 43.1% of total production in West Africa and Africa, respectively . With an estimated yearly production of 5.49 million metric tonnes, Nigeria produces almost 37% of the world's total, with Ghana producing 31% . However (Eze and Okorji, 2003) reported that Nigeria produced almost 40% of the world's cocoyam; this suggests that the country's cocoyam crop is diminishing (Zuhair and Hunter, 2000). Sand loamy soil that drains effectively is ideal for cocoyam growth. When grown in rich soil with a high capacity for water retention, it delivers the maximum amount of fruit (Lewu *et al*., 2010). Planting is typically done at or just before the start of the rainy season in the wet tropics when irrigation is not available. It grows by using setts and takes 8 to 12 months to reach maturity after planting.

It is believed that cocoyam possesses more nutritional value than other major West African roots and tubers, particularly when it comes to protein digestibility and mineral composition (magnesium, phosphorus, and calcium) (Lim, 2016). It can be chopped up and boiled or fried to form crispy chips or flakes, or it can be used as a thickening in soup or flour. You can consume the leaf stalk as well. In many underdeveloped nations, cocoyams account for a sizable amount of the diet's carbohydrate content and produce edible starchy storage corms and cormels. However, it has been claimed that cocoyam contains oxalate, which when consumed in meals made from them might cause discomfort, toxicity, or both (Quash, 2000). Corms can be stored at room temperature for a respectable amount of time, but for long-term preservation, they should be kept at 7oC. Improper treatment of the cocoyam after harvest or storage is the cause of post-harvest deterioration.

The most important element in cocoyam spoiling is thought to be pathogen invasion through natural holes or wounds (Osuji, 2003). But because of the challenges with storage, cocoyams are typically used or eaten fresh soon after harvest because fungus cause them to decay while being stored . *Aspergillus flavus, Fusarium oxysporum, Penicillium cyclopium, Penicillium digitatum, Aspergillus niger, Botryodiplodia theobromae, Sclerotia rolfsii, Mucor circinelloides, Rhizopus stolonifer, Fusarium solani*, and the bacteria *Erwinia carotovora* are among the common species of microorganisms associated with cocoyam rot in Nigeria (Agu et al., 2016). Fungi damage cocoyams by colonizing them and depolymerizing particular cell wall polymers. Given that cocoyams are a food plant that is widely consumed, handling, planting, storing, cooking, and even preservation of the plant should be done with extreme caution. It is important to take aseptic precautions, such as using clean planting tools, to completely prevent fungal growth, which can endanger consumer health. Therefore, ensuring the safety of this food product is of utmost importance and calls for everyone's participation in the hand-to-fork chain. This made it necessary for this study to look into the fungi that cause cocoyam rotting.

**MATERIALS AND METHODS**

 **Collection of Materials**

We took cormels of cocoyam (*Colocasia esculenta linn*) that showed signs of post-harvest rot from a cocoyam barn in a neighborhood market in Benin City, Nigeria. From the same barn, fresh, healthy cocoyam cormels were also gathered. The cocoyam corms were moved to the lab for additional analysis.

 **Isolation**

The spoiled tissues were removed from the cocoyam corms using a sterile knife and forceps. Using a surface-sterilized forcep, 2 mm cubes were removed from the tissue at the intersection of the healthy and infected portions of the cocoyam cormels. They were cleaned twice (one minute each wash) with sterile distilled water after being surface sterilized (to get rid of surface impurities) in 70% ethanol. The cormel piece was dried for ten minutes in a Laminar Air flow Hood chamber by being placed on sterile paper towels. According to the procedure used by (Agu et al., 2016), a portion of the spoilt area of the cocoyam was cut out with a sterile knife, and the rotten parts were pushed into the center of the agar plate as an inoculum on Potato dextrose agar (PDA), which contains gentamicin, an antibacterial agent. Following the inoculation, plates were incubated for four days at 27°C, and every day they were checked to see if any fungal growth had developed. Repeated subculture techniques were used to purify the growing cultures. Subculturing, Identifying, and Characterizing Test Fungi Pathogens

Once growth was achieved, subcultures were made using inocula from the different species in the mixed cultures. Using flame-sterilized blades, hyphal tips were transferred from the colony edge of the mixed cultures to new PDA plates that contained gentamicin, an antibacterial agent. Until pure cultures were established, plates were subcultured at 27°C in an incubator. The Petri dishes holding the pure cultures of the test fungi were then sealed with parafin to avoid contamination. The fungal colonies' gross morphological appearance on PDA culture media was described in the Manual of Fungal Atlas, which guided the characterisation and identification of the isolates and the method of slide culture for microscopic analysis (Wagner et al., 1999)

 **Test for Pathogenicity**

For the pathogenicity test, methodology described by (Okigbo, 2003) was used. The pathogenicity test was then conducted using the test fungal isolates that were subcultured from the spoiled cocoyam corms. After thoroughly cleaning with tap water and rinsing with distilled water, healthy cocoyam corms were used. The corms' surfaces were then cleaned of infection using a 70% ethanol solution. The cormels were put in a Laminar Air flow hood and let to dry for 12 minutes on sterile paper towels. Two distinct holes were drilled into the cocoyam corms using a sterile cork borer with a 5 mm diameter. Pure fungal isolates were then added to the holes using a second sterile cork borer, and the holes were sealed with petroleum jelly. One milliliter of sterile distilled water was used to inoculate controls who were not exposed to any isolate. Following the complete inoculation of the test isolates into their corresponding healthy corms, each corm was incubated and monitored in a humidity chamber for a duration of 14 days. Every day, the corms were inspected for signs of discoloration, shrinkage, and loss of weight. The weight reduction as a percentage aims to quantify the extent of the infection and the pathogenicity rate of the fungi that cause rot.

This was ascertained by weighing each individual yam tuber at completion and comparing it to its starting weight before inoculation. After 14 days, the weight loss of the cocoyam corms measured as a percentage.

IW-FW x 100 = % weight loss in IW, where FW = Final weight of infected cocoyam tuber after 14 days

IW stands for initial tuber weight before to inoculation.

**RESULTS AND DISCUSSION**

The purpose of this study was to assess the fungal pollutants linked to the spoiling of cocoyam that was purchased in Benin Metropolis, Edo State, Nigeria

In Edo State's New Benin, Benin City, a local barn provided the cocoyam used for the study. It was discovered that these cocoyam corms were infected by fungi, and these fungi were isolated using potato dextrose agar, a growth medium that contains the antibiotic drug gentamicin. *Cercinelloides, Aspergillus niger, Aspergillus fumigatus, Rhizopus stolonifer, Fusarium oxysporum,* and *Penicillium cyclopium* were the fungus found in this damaged cocoyam. After inoculating samples of healthy cocoyams with the identified fungal species, the samples were monitored for alterations and spoiling over a period of 14 days.

The following summarizes the findings of the analysis of the fungal pollutants linked to the spoiling of cocoyam samples

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| **Table 1: Colonial morphology and microscopic characterizations of fungi associated with cocoyam spoilage.** |
| No. | Colonial characteristics | Microscopic features | Rot type  | Probable fungi |
| i.  | Colonies were fluffy and cottony, and had a yellow reverse side. There was rapid growth and the plate was completely covered with growth within 3 days. | hyaline, non-septate, branching and sympodially and circinate were the characteristics of sporangiophores. No chlamydospores. | Soft | *M. cercinelloides* |
| ii. | Fast growth rate. Colonies that are compact and level with a yellow basal foot. Pale yellow on the back | septate hyphae with soft , simple, erect pale brown conidiophores with foot cells at the base.  | Soft | *Aspergillus niger* |
| iii. | White-black colonies that expanded quickly. White on the back side. | Hyphae had green conidiospores and was septate. basal foot cell presence.  | Soft | *Aspergillus fumigatus* |
| iv. | swift expansion of colonies, cottony and turn yellow, then dark grey, then back to white.   | Grey, smooth-walled sporangiospores feature massive, globose sporangia and well-developed subtending columellae. | Dry | *Rhizopus stolonifer* |
| v. | A pink, rapidly expanding cottony colony | The largest spores were packed with cells and resembled a sickle. | Soft | *Fusarium oxysporum* |
| vi. | The colonies are velvety, green, and growing quickly. | Conidia has a smooth, ellipsoidal shape. Mycelia arranged erratically.  | Dry | *Penicillium cyclopium* |

The microscopic characterizations and colonial morphology of the fungi linked to cocoyam rotting are listed in Table 1. Mucor cercinelloides, Aspergillus niger, Aspergillus fumigatus, Rhizopus stolonifer, Fusarium oxysporum, and Penicillium cyclopium were the six fungal species that were isolated from the spoiled cocoyam cormels.

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| **Table 2: Characteristic observations in the inoculated cocoyam samples observed for 14 days** |  |
| Days | *M. cercinelloides* | 1. *Niger*
 | 1. *fumigatus*
 | *R. stolonifer* | *F. oxysporum* | *P.* cyclopium |
| 1 | unchanged | Unchanged | Unchanged | Unchanged | Unchanged | Unchanged |
| 2 | Unchanged | Unchanged | Unchanged | Unchanged | Unchanged | Unchanged |
| 3 | Unchanged | Unchanged | Unchanged | Unchanged | Unchanged | Unchanged |
| 4 | Tiny whitish mucoid material | filamentous substance and black in color observed |  dust-like growth and greenish in color observed | Greyish substance with yellowish dusts  | Bud-like whitish substance observed | Unchanged |
| 5 | More visible white mucoid substance | More observable filamentous blackish substance | Greenish substance became more visible | Greyish substance became more observable | More white buds seen | Unchanged |
| 6 | A white mucoid substance spraed on the sample | blackish filaments formed a cotton-like wool | Continued dissemination of the dust-like greenish substance | Sample became darker in colour | white bud-like substance was visible and spreading throughout  | Unchanged |
| 7 | Sample began to shrink | Sample began to shrink | Sample contracted and colour darkened | Continued dispersal of the greyish substance  | Weight began to decrease | Unchanged |
| 8 | Significant weight loss | Significant weight loss | Significant weight loss | Reduction in size | Noticeable color dullness and shrinkage | Whitish substance seen in a single location |
| 9 | Sample dried up and lost weight | Sample appeared dry and shrunk | Sample appeared dry | Significant weight reduction observed | Further shrinking and dryness | No spread of dot-like whitish substance |
| 10 | No further alterations | No further alterations | No further alterations | No further alterations | No further alterations | No further alterations |
| 11 | No further alterations | No further alterations | No further alterations | No further alterations | No further alterations | Fair shrink in size |
| 12 | No further alterations | No additional changes | No further alterations | No further alterations | No further alterations | Weight reduction observed |
| 13 | No further alterations | No further alterations | No further alterations | No further alterations | No further alterations | No further alterations |
| 14 | No additional changes | No additional changes | No additional changes | No further alterations | No further alterations | No additional changes |

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| The distinctive findings in the infected cocoyam samples that were monitored for 14 days are displayed in Table 2. The six fungal species listed and reported in Table 1 were injected into the cocoyam samples.  The weight of the cocoyam before and after the fungus species inoculation is displayed in Table 3. Between 49.57% and 64.66% of the cocoyam samples that were injected with various fungal infections showed a percentage drop in weight**Table 3: Weight of cocoyam before and after inoculation with the fungal species** |
| Fungi | Before (g) | After (g) | Percentage weight reduction (%) |
| *M. cercinelloides* | 71.5 | 30.0 | 58.04 |
| 1. *niger*
 | 107.0 | 45.0 | 57.94 |
| 1. *fumigatus*
 | 84.9 | 30.0 | 64.66 |
| *R. stolonifer* | 95.0 | 48.0 | 49.47 |
| *F. oxysporum* | 84.9 | 36.8 | 56.65 |
| *P. cyclopium* | 96.2 | 46.3 | 51.87 |
| **Control** | 53.0g | 51.00g | 3.77 |

With the exception of *Aspergillus fumigat*us, all six fungal species recovered from the current samples were also isolated by (Agu et al., 2016), whose study is consistent with the fungal species isolated in this investigation. (Onuegbu, 1999) examined the root and related fungal infections in cocoyam and identified *Aspergillus niger, Botryodiplodia spp., Corticium rolfsii, Geotrichum candidum, and Fusarium spp*. as rot-causing fungi. These fungi, in particular, Fusarium and Aspergillus, were also identified way back. (Ugwuanyi, 1996) reported that the Aspergillus rot was so severe that the cocoyam tissues completely macerated. Four of these six fungi—*Aspergillus niger, Penicillium, Rhizopus stolonifer,* and *Fusarium spp*.—were also isolated by Ankworji, (2012) during the study of fungi that cause cocoyam rot. According to Frank and Kingsley, (2014) these organisms are true crop diseases that affect root and tuber crops. According to (Okigbo, 2003) the Rhizopus and Mucor that were isolated belonged to the fast-growing fungal species that are thought to be responsible for the cocoyam rot.

No discernible change was seen until day 3 for all of the samples infected with different fungal species, as shown in Table 2. Conversely, the Penicillium-inoculated sample did not exhibit any notable alterations until the eighth day of observation. Additionally, it was noted that, after day 9, no additional alterations were seen in the cocoyam samples infected with the different fungal infections (with the exception of *Penicillium cyclopium*, which showed shrinkage and weight decrease up to day 12). Perhaps the cormel's continued dramatic modifications when others had stopped could be attributed to the delayed expression of *Penicillium cyclopium* infection.

As a result of the fungal pathogens' pathogenicity, the percentage weight loss varied from 49.47% to 64.66%, while it was only 3.77% in the unvaccinated controls. One possible explanation for the 3.77% could be the typical dehydration that comes with storage. Nevertheless, the cause of a percentage loss as high as 64.66% is not simply dehydration; rather, it is the result of the infection with these fungi. Should this food product be contaminated by such organisms, producers and customers may suffer significant financial losses because to the dramatic decrease in cocoyam weight over such a short period of time. This is consistent with report by Osuji, (2003) who concluded that fungal diseases are the main cause of the over 40% of cocoyam that may be lost to post-harvest rot. In this investigation, every organism significantly changed the cormels' size, morphological characteristics, and color. Particularly in the barns, these tubers are frequently kept for extended periods of time in storage. The identification of numerous pathogenic fungi from a single cormel indicates the potential for many infections, which when combined, might quickly lead to the rotting of crops that have roots and tubers. According to (Agu et al., 2016) because of variables including temperature and relative humidity, these fungi have been discovered to generate damaging rot blight complex (CRRBC), which poses a serious danger to cocoyam production.

The fungus infect this crop prior to harvest, but they also cause harm after harvest due to improper handling, insects, and other animal damage, as well as through direct encroachment of these pathogenic organisms into the plant's undamaged epidermis. However, all of these can be prevented by using hygienic planting supplies and equipment, as well as by properly packaging and managing them.

**CONCLUSION**

Cocoyams should be planted, stored, cooked, and even preserved with extreme care and caution because they are a food plant that is widely consumed. Aseptic measures are necessary to totally prevent fungal growth, which can endanger consumers. Examples of these precautions include the use of clean planting tools. The spread of knowledge about appropriate planting methods and considerations for growing cocoyam and tubers in general is necessary in order to foster a healthy country and a balanced economy.

**ETHICS**

 The study proposal was presented and then approved by the Research committee of the Department of Microbiology, University of Benin, Nigeria.

**AUTHOR DISCLOSURE STATEMENT**

The authors declare no conflict of interest.

Anderson JW, Baird P, Davis RH JR, Ferreri S, Knudtson M, Koraym A, Waters V, Williams CL. (2009). Health benefits of dietary fiber. Nutrition Reviews. 2009. 67(4):188-205.

**REFERENCES**

Agu KC, Awah NS, Nnadozie AC, Okeke BC, Orji MU, Iloanusi CA, Anaukwu CG, Eneite HC, Ifediegwu MC, Umeoduagu ND, Udoh EE. (2016). Isolation, identification and pathogenicity of fungi associated with cocoyam (*Colocasia esculenta*) spoilage. Food Science and Technology*.* 4(5): 103-106.

Anukworji CA, Putheti RR, Okigbo RN. (2012). Isolation of fungi causing rot of cocoyam (*Colocasia esculenta* (l.) schott) and control with plant extracts: (*Allium sativum*, l., garcinia kola, heckel., *Azadirachta indica*, l.and *Carica papaya*, l.). Global Advanced Research Journal of Agricultural Science1(1): 1-15.

 Eze CC, Okorji EC. (2003). Cocoyam production by women farmers under improved and local technologies in Imo State, Nigeria. Nigeria journal of Science1: 133-166.

Frank CO, Kingsley CA. (2014). Proximate composition, physiological changes during storage, and shelf life of some Nigerian varieties of yams (*Dioscorea* species). Journal of Scientific Research and Reports 3(4): 553-562.

Lewu MN, Adebola PO, Afolayan AJ. (2010). Comparative assessment of the nutritional value of commercially available cocoyam and potato tubers in South Africa. Journal of Food Quality 33**:** 461-476.

 Lim TK. (2016). *Edible medicinal and non-medicinal plants.*1st edn. New York, London: Springer. https://link.springer.com/book/10.1007/978-94-007-1764-0

Okigbo RN. (2003). Fungi associated with peels of post-harvest yam in storage. Global Journal of Pure and Applied Science 9: 19-23.

Onuegbu BA. (1999). Evaluation of the efficacy of fungicides, plant extracts and chemicals in minimizing mould growth in mung bean (*Vigna radiata L*.) seeds. Legume Research, 22(4): 270-272.

 Osuji JO. (2003). Cytogenetic techniques. In E.N. Onyeike and J.O. Osuji (Eds.) Research Techniques in Biology and Chemical Science*,* 70-83.

 Quash ML, Melton LD, Harris PJ, Burdon JN, Smith BG. (2000). Cell wall compositions of raw and cooked corms of taro (Colocasia esculenta). Journal of Science Food and Agriculture 81: 311-318.

Ugwuanyi JO. (1996). *Mycopathologia*. Department of Microbiology University of Nigeria, Nsukka, Nigeria.

Wagner WL, Herbst DR, Sohmer, SH. (1999). Manual of the flowering plants of Hawai‘i. Revised edition, 2, University of Hawaii Press/Bishop Museum Press.

 Zuhair M, Hunter DG (2000). Taro cultivation and use in the Maldwes. IP*GR* Session 12th Symposium of IATRC,TSUKUBA. Japan.